



## Tumors of salivary glands: an institutional study

Kanala Mahaboob Chan<sup>1</sup>, P Chaturya Kalanidhi<sup>2</sup>, Vaddadi Vishali<sup>3</sup>, Muchalapuri Sravani<sup>4\*</sup>, Rasheed Fatima<sup>5</sup>, Balagouni Shruthi<sup>6</sup>, Sumanjali Chari<sup>6</sup>

<sup>1</sup>Assistant Professor, Department of Pathology, Government Medical College, Ongole, Andhra Pradesh

<sup>2</sup>Senior Resident, Department of Pathology, Government Medical College, Nalgonda

<sup>3</sup>Associate Professor, Department of Pathology, Khaja bandanawaz University, Faculty of Health Sciences, Kalaburgi

<sup>4</sup>Assistant Professor, Department of Pathology, SVS Medical College and Hospital, Yenugonda, Mahabubnagar, Telengana

<sup>5</sup>Professor, Department of Pathology, SVS Medical College and Hospital, Yenugonda, Mahabubnagar, Telengana

<sup>6</sup>Postgraduate, Department of Pathology, SVS Medical College and Hospital, Yenugonda, Mahabubnagar, Telengana

**\*Corresponding author:** Muchalapuri Sravani, **Address:** Department of Pathology, SVS Medical College and Hospital, Yenugonda, Mahabubnagar, Telengana-509001, **Email:** sravani.muchalapuri@gmail.com **Tel:** +9185422 31188

### Abstract

**Background & Aims:** This research investigates the clinicopathological features of salivary gland tumors by employing a range of markers, such as pancytokeratin (CKAE1/CKAE3),  $\alpha$ -smooth muscle actin (SMA), carcinoembryonic antigen (CEA), and p63.

**Materials & Methods:** The research investigated formalin-fixed paraffin-embedded (FFPE) tissue slides stained with hematoxylin and eosin (H & E) from a total of 32 cases of salivary gland tumors.

**Results:** Lesions were detected in individuals ranging from 11 to 79 years of age, with a mean age of 45.5 years. Among the cases analyzed, 19 (59.4%) were categorized as benign neoplasms, which included 12 instances of pleomorphic adenoma (PA), 4 instances of basal cell adenoma (BCA), 2 instances of myoepithelioma, and 1 instance of Warthin's tumor. Conversely, 13 cases (40.6%) were identified as malignant neoplasms, consisting of 7 cases of mucoepidermoid carcinoma, 4 cases of adenoid cystic carcinoma (ACC), 1 case of carcinoma ex pleomorphic adenoma (Ca ex PA), and 1 case of squamous cell carcinoma (SCC). Immunohistochemical analysis demonstrated p63 positivity in PA, BCA, ACC, and SCC. Furthermore, positivity for cytokeratin was found in BCA, ACC, PA, and SCC. SMA positivity was observed in both BCA and ACC.

**Conclusion:** The predominant benign and malignant tumors of the salivary glands are PA and mucoepidermoid carcinoma (MEC). Immunohistochemical staining of both epithelial and myoepithelial cells serves not only to validate the diagnosis but also assists in formulating the most appropriate treatment strategy.

**Keywords:**  $\alpha$ -smooth muscle actin, Carcinoembryonic antigen, Salivary gland tumors, Squamous cell carcinoma

Received 21 October 2024; accepted for publication 15 March 2025

### Introduction

Salivary gland neoplasms account for 3-10% of all tumors identified in the head and neck region (1). A

variety of bioactive proteins have been discovered in saliva (2-4). The salivary glands play a crucial role in the production of immunoregulatory and anti-inflammatory agents (5-7).

These glands are classified into three pairs: the parotid, submandibular, and sublingual glands, in addition to numerous minor glands that are distributed throughout the area. The minor salivary glands are particularly numerous, with estimates suggesting there are between 600 and 1,000 clusters located within the oral submucosa, excluding the anterior hard palate and gingiva (8, 9). They are also present in the submucosal layers of the paranasal sinuses, pharynx, larynx, trachea, bronchi, lips, palate, and oropharynx. Moreover, heterotopic minor salivary glands can form in unusual locations such as lymph nodes, the thyroid gland capsule, facial bones, and the pituitary gland. Approximately 12% of all salivary gland tumors originate from the lachrymal and mucous glands situated on the inner surface of the upper lip, palate, base, and margins of the tongue, nasopharynx, larynx, and accessory nasal sinuses (10).

Salivary gland neoplasms are relatively rare, representing less than 1% of all cancers, with a significant proportion of these tumors being benign (11, 12). These neoplasms can arise in both major and minor salivary glands. The parotid glands are involved in 80% of cases, the submandibular glands in 10%, while the remaining cases are attributed to the sublingual and various minor salivary glands (13, 14). Tumors in the major salivary glands are frequently benign, whereas those in the minor salivary glands are more likely to be malignant. The malignancy rate for tumors in the parotid gland ranges from 20% to 25%, increases to 40% for submandibular gland tumors, and exceeds 90% for sublingual gland tumors (15,16). Additionally, benign salivary gland tumors are more commonly found in females than in males (17). The parotid gland is the most prevalent location for salivary gland tumors, accounting for 70% of all cases (18, 19). Within this category, PA is the most common benign tumor, representing 80% of occurrences, followed by Warthin's tumor (20). In most studies, mucoepidermoid carcinoma (MEC) and adenoid cystic carcinoma (ACC) are identified as the most frequently encountered malignant tumors.

Although hematoxylin-eosin (H&E) staining is the conventional technique for the identification of salivary

gland tumors, the use of immunohistochemistry (IHC) can improve diagnostic accuracy, despite certain limitations in its effectiveness.

The anatomy of the salivary gland consists of ducts and acini (ducto-acinar units), which are made up of four distinct cell types: ductal, acinar, myoepithelial, and basal cells. Pan-cytokeratin (CKAE1/CKAE3) is present in all four cell types, while epithelial membrane antigen (EMA) and carcinoembryonic antigen (CEA) are expressed in both ductal and acinar cells. Myoepithelial and basal cells show positivity for CK14 and p63, but are negative for EMA and CEA. Only myoepithelial cells express markers such as smooth muscle actin (SMA), muscle-specific actin (MSA), podoplanin, calponin, and vimentin. The staining pattern for S-100 protein differs among the four cell types.

This study aims to explore the clinicopathological features of salivary gland tumors and their relationship with immunohistochemical markers, including CKAE1/CKAE3, CEA, SMA, and p63.

## Materials & Methods

This retrospective study was conducted from April 2019 to April 2022. Case information was acquired from the department's histopathological records from 32 total cases. Inclusion criteria comprised all cases of salivary gland neoplasms. Inflammatory lesions and non-neoplastic cysts were considered as exclusion criteria.

Aspiration was performed using 22-gauge needles and smears were fixed with 95% ethyl alcohol. Formalin-fixed and paraffin-embedded (FFPE) sections were stained with H&E. IHC was performed using markers including CKAE1/CKAE3, CEA, SMA, and p63.

## Results

Clinical symptoms included sensations of numbness, muscle weakness, and persistent pain localized to the salivary gland region. Salivary gland lesions were observed in individuals aged between 11 and 79 years, with a mean age of 45.5 years. The cohort consisted of 14 male patients (46.67%) and 16 female patients

(53.33%). A higher incidence of malignant lesions was found among male patients, while female patients presented a greater number of benign lesions.

Among the total cases, 13 (40.6%) were categorized

as malignant neoplasms, which comprised 7 cases of mucoepidermoid carcinoma, 4 cases of ACC, one case of carcinoma ex pleomorphic adenoma (Ca ex PA), and one case of squamous cell carcinoma (SCC) (Table 1).

**Table 1.** Basic characteristic features of salivary gland tumors

	No. of cases	Percentage
<b>Tumors</b>		
Benign	19	59.4%
Malignant	13	40.6%
<b>Benign tumors</b>		
PA	12	63.15%
Basal cell adenoma (BCA)	04	21.06%
Myoepithelioma	02	10.53%
Warthin's tumor	01	5.26%
<b>Malignant tumors</b>		
MEC	07	53.85%
ACC	04	30.77%
Ca ex PA	01	7.6%
SCC	01	7.6%
<b>Age incidence</b>		
< 30 yr	7	23.33%
30-39 yr	5	16.67%
40-49 yr	3	10%
50-59 yr	4	13.33%
60-69 yr	5	16.67%
70-79 yr	6	20%
<b>Sex</b>		
Female	16	53.33%
Male	14	46.67%
<b>Sites</b>		
Parotid gland	18	60.00%
Submandibular gland	7	23.33%
Sublingual salivary gland	3	10.00%
Minor salivary gland	2	6.67%

Table 2 depicts the distribution of salivary gland tumors according to their specific anatomical locations.

Table 3 offers a comprehensive overview of the site-specific distribution of malignant salivary gland tumors.

**Table 2.** Site-wise distribution of benign salivary gland tumors neoplasms

Neoplasms	Parotid	Submandibular	Sublingual	Minor salivary gland	P-value
PA	07	03	02	0	0.89*
BCA	04	0	0	0	
Myoepithelioma	02	0	0	0	

Neoplasms	Parotid	Submandibular	Sublingual	Minor salivary gland	<i>P-value</i>
Warthin's tumor	01	0	0	0	
Total	14	03	02	00	

\*Chi-Square test

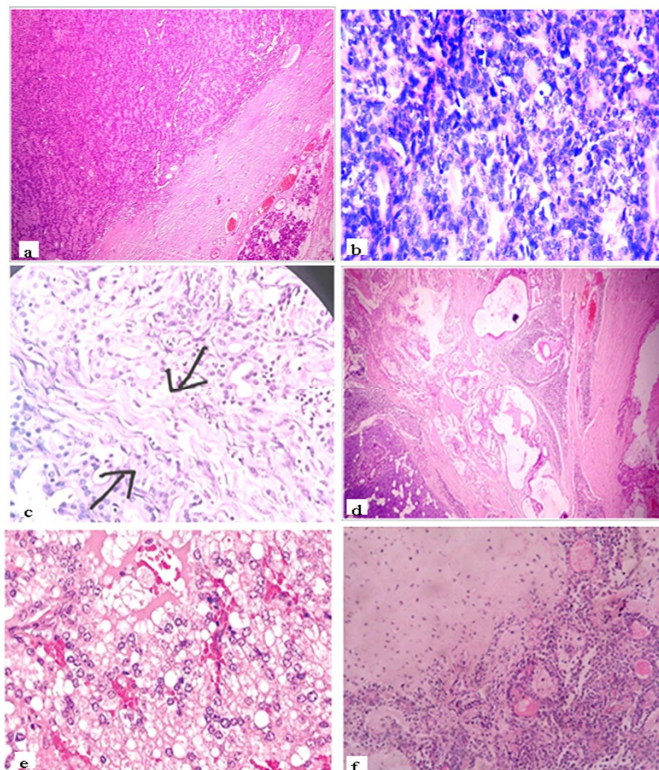
**Table 3.** Site-wise distribution of malignant salivary gland tumors

Malignant	Parotid	Submandibular	Sublingual	Minor salivary gland	<i>P-value</i>
MEC	05	01	0	01	0.85*
ACC	02	01	0	01	
Ca ex PA	01	0	0	0	
SCC	01	0	0	0	
Total	09	02	0	02	

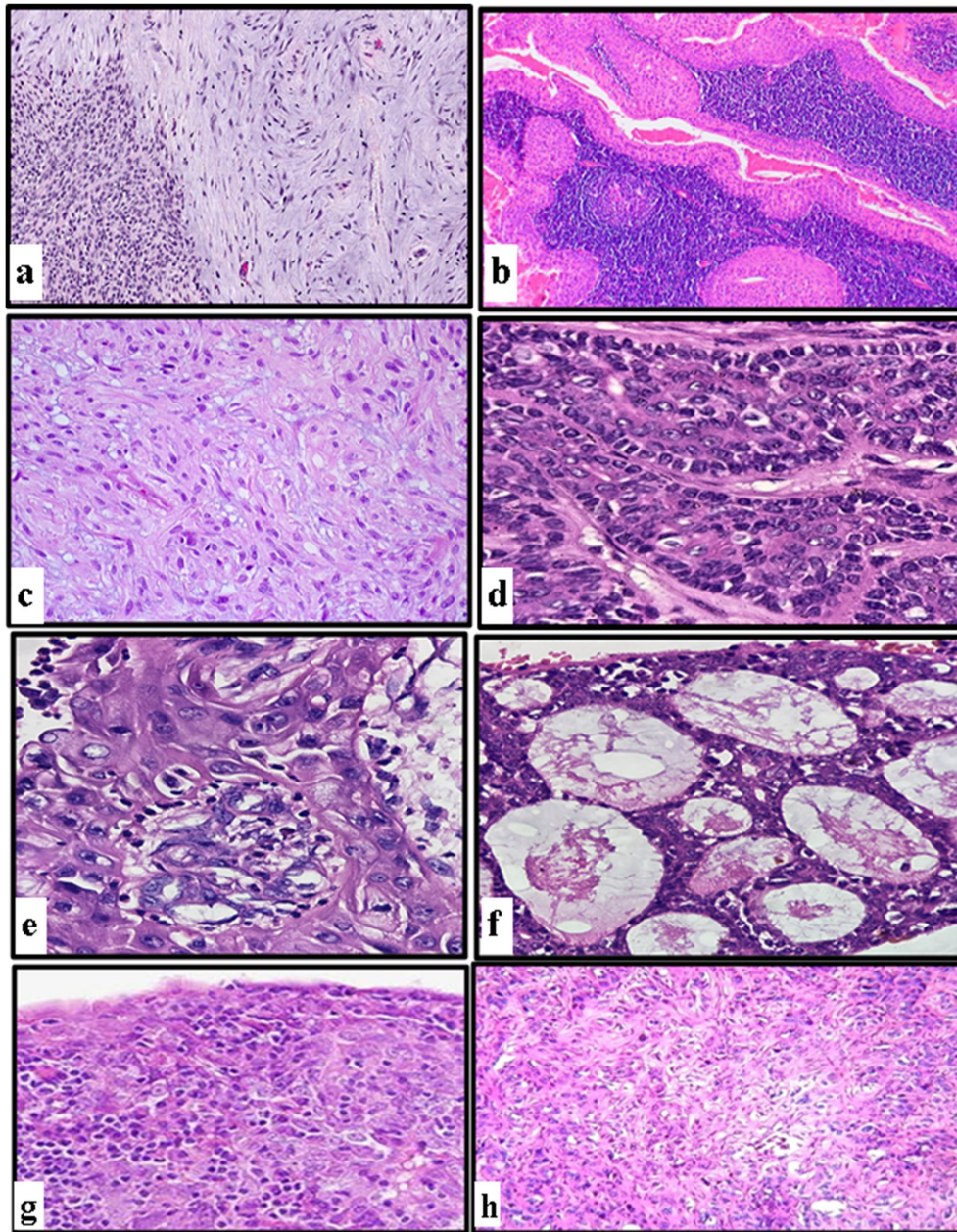
\*Chi-Square test

Figure 1 displays H&E staining results for various tumors, including BCA, ACC, MEC featuring anaplastic epidermal cells and mucus-producing cells, acinic cell carcinoma, and PA, highlighting a variety of histological

characteristics. Figure 2 further illustrates PA, Warthin's tumor with papillary structures, spindle-shaped myoepithelial cells, BCA, mucoepidermoid carcinoma, ACC, SCC, and Ca ex PA, emphasizing their diverse features.

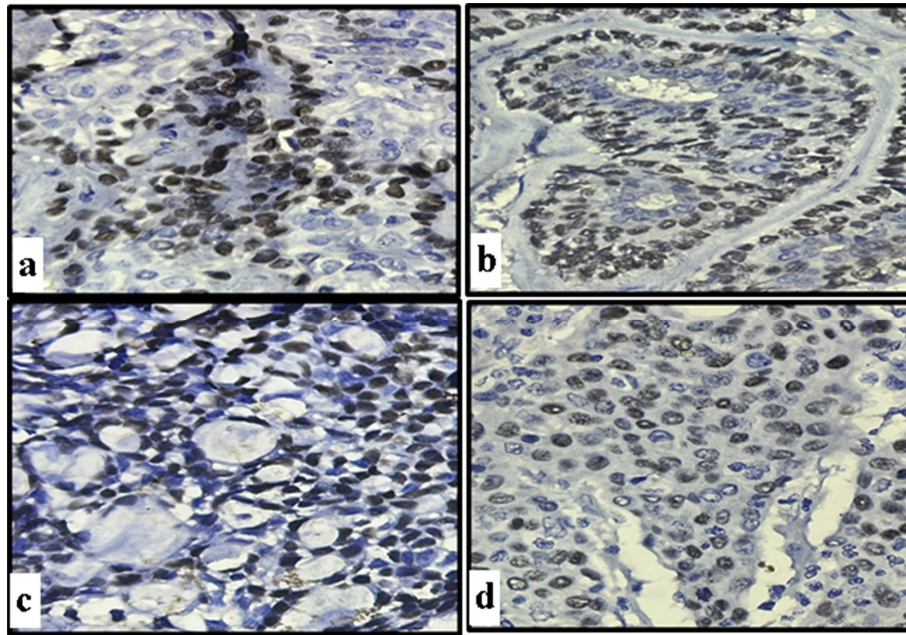


**Fig. 1.** **a.** BCA with adjacent normal parotid gland (×100). **b.** BCA with trabecular type (×400). **c.** ACC with perineural invasion (×400). **d.** MEC with epidermal cell anaplasia & mucus cells (×100). **e.** Acinic cell carcinoma (×400). **f.** PA with chondroid differentiation (×100).



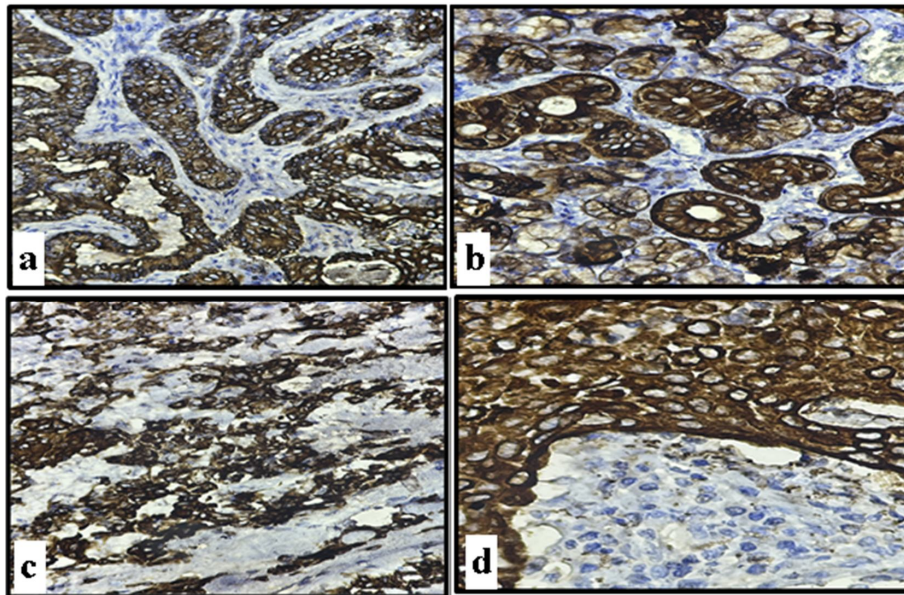
**Fig. 2.** H & E staining. **a.** PA - Spindle-shaped myoepithelial x-cells in a myxoid background. **b.** Warthin's tumor- Papillary structures lined by bilayered oncocytic epithelial cells, surrounded by a lymphoid stroma. **c.** Myoepithelioma- Spindle shaped myoepithelial cells. **d.** BCA- Solid tumor nests with a peripheral palisading manner. **e.** Mucoepidermoid carcinoma- Showing epidermal, mucin and intermediate cells. **f.** ACC- Cribriform pattern is composed predominantly of myoepithelial cells, admixed with hyalinized or myxoid globules. **g.** SCC- Showing pleomorphic cells. **h.** Ca ex PA- Showing both benign and malignant components. Magnification:  $\times 400$ .

Figure 3 presents the findings of an immunohistochemical analysis indicating p63 positivity in PA, BCA, ACC, and SCC.



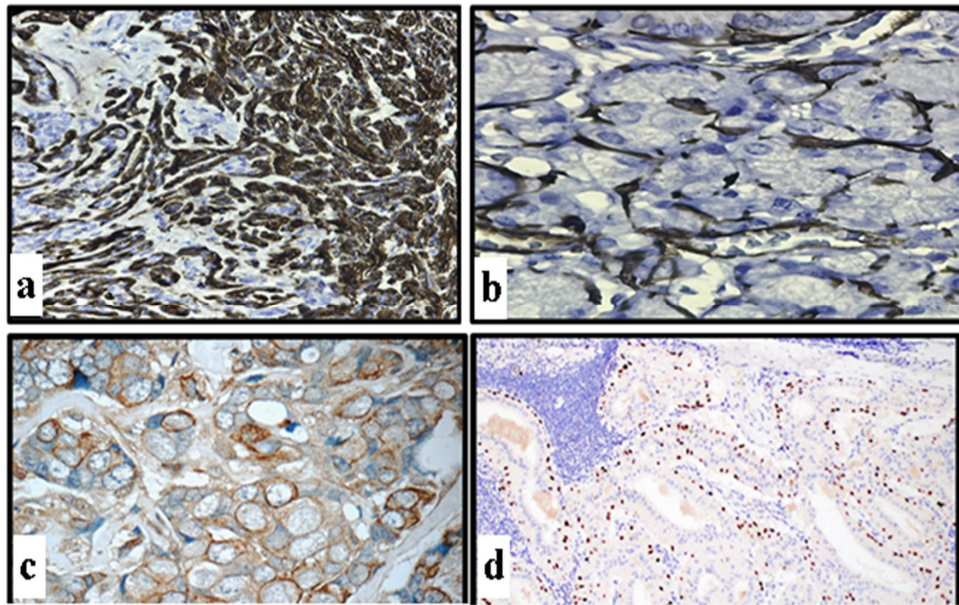
**Fig. 3.** IHC: p63 Positivity. **a.** PA (Myoepithelial cells + ve for p63 & SMA). **b.** BCA (basaloid cells + ve for p63). **c.** ACC (Myoepithelial component is positive for SMA, p63, GFAP & cytokeratins). **d.** SCC (P63 + ve for basaloid type cells).

Figure 4 illustrates the immunohistochemical evaluation of cytokeratin positivity in BCA, ACC, PA, and SCC.



**Fig. 4.** IHC: Cytokeratin Positivity. **a.** BCA. **b.** ACC. **c.** PA. **d.** SCC

Finally, Figure 5 showcases the immunohistochemical assessment of SMA positivity in BCA and ACC.



**Fig. 5.** IHC: SMA positivity. **a.** BCA. **b.** ACC. **c.** Cytokeratin positive in mucoepidermoid carcinoma. **d.** P63 in Warthin's tumor.

## Discussion

Salivary gland tumors occur at an estimated rate of 0.4 to 13 cases per million individuals each year, with 70 to 80 percent of these tumors arising in the parotid gland. Fine needle aspiration cytology (FNAC) plays a crucial role in distinguishing between benign and malignant lesions. In the present study, the mean age of participants was 45.5 years, with ages ranging from 11 to 70 years. Comparable results have been documented in the studies conducted by Shrestha et al. and Bashir et al (21, 22).

Our findings revealed a higher incidence of tumors in females than in males. Specifically, female patients showed a greater prevalence of benign lesions, whereas males were more often diagnosed with malignant tumors. The parotid gland emerged as the most frequently affected site, representing 60% of the cases (18 cases), followed by the submandibular gland at 23.33% (7 cases) and the sublingual salivary gland at 10% (3 cases). Minor salivary glands were implicated in two cases, accounting for 6.67%.

The predominance of parotid gland involvement observed in our study is consistent with the findings of Bashir et al. (22), Cohen et al. (23), and Chatterjee and Panda (24). Among the 32 cases examined, 19 (59.4%) were classified as benign neoplasms, which included 12 cases of PA, 4 cases of BCA, 2 cases of myoepithelioma, and 1 case of Warthin's tumor. In contrast, 13 cases (40.6%) were identified as malignant neoplasms, consisting of 7 cases of mucoepidermoid carcinoma, 4 cases of ACC, 1 case of Ca ex PA, and 1 case of SCC.

According to Bhargava et al. (25), the most common malignant tumor is ACC (14%), followed by MEC (3%) and acinic cell carcinoma (2%). In our analysis, histopathological evaluation indicated that benign tumors were predominantly PAs (59.4%), while malignant tumors were primarily mucoepidermoid carcinomas (40.6%). The higher prevalence of benign neoplasms relative to malignant ones in our study aligns with findings from prior research (26, 27, 28).

In our study, PA was identified as the most common benign tumor, accounting for 63.15% of the cases. This observation is consistent with findings from prior

research. The current results confirm that PA is the leading benign neoplasm, followed by monomorphic adenoma, supporting earlier studies. Furthermore, our analysis indicated that 53.85% of the cases were classified as MEC, aligning with the results reported by Panda and Agarwal (5%), 30. Khazanchi et al. (9.5%), and Soni et al. (8%) (29, 30, 31).

Among the malignant tumors, MEC was found to be the most prevalent, followed by ACC and acinic cell carcinoma. The identification of low-grade MEC presents considerable diagnostic challenges, as it may be incorrectly diagnosed as chronic sialadenitis, Warthin's tumor, mucous retention cysts, or mucous salivary gland adenomatoid hyperplasia. Diagnosis is established through H & E staining, followed by confirmation with IHC.

Immunohistochemical analysis demonstrated that epithelial cells express various markers, including cytokeratins, EMA, and carcinoembryonic antigen (CEA) (32, 33). In contrast, myoepithelial cells do not show epithelial markers but express specific myoepithelial markers such as p63 and SMA. In our research, p63 was used to identify abluminal myoepithelial cells, as it is recognized as the most reliable myoepithelial marker in these tumors (34, 35). The immunohistochemical findings indicated that the clear abluminal cells are of myoepithelial origin, while the luminal cells, characterized by eosinophilic cytoplasm, are derived from ductal cells.

### Conclusion

The most commonly observed benign and malignant tumors of the salivary glands are PA and mucoepidermoid carcinoma, respectively. Histopathological analysis is considered the definitive method for diagnosing salivary gland tumors. The use of immunohistochemical staining on epithelial and myoepithelial cells plays a crucial role in verifying the diagnosis, which in turn aids in determining an appropriate treatment strategy.

### Acknowledgments

The authors acknowledged the lab staff for allowing the use of the departmental laboratory and other necessary facilities needed.

### Ethical statement

The proposal for this study was approved by the Ethics Committee of SVS Medical College, Mahabubnagar (IEC/SVS/2023/Pathology). The researchers adhered to all the principles recommended by the Helsinki Convention on Ethics in Research. Patients' personal and attributable information were kept confidential.

### Data availability

The data that support the findings of this study are available upon request from the corresponding author.

### Conflict of interest

The authors declare no conflict of interest.

### Funding/support

No funding support was received for this study.

### Author contributions

None declared.

### References

1. Ansari MH. Salivary gland tumors in an Iranian population: a retrospective study of 130 cases. *J Oral Maxillofac Surg* 2007;65(11):2187-94. <https://doi.org/10.1016/j.joms.2006.11.025>
2. Pedersen AM, Bardow A, Jensen SB, Nauntofte B. Saliva and gastrointestinal functions of taste, mastication, swallowing and digestion. *Oral Dis* 2002;8(3):117-29. <https://doi.org/10.1034/j.1601-0825.2002.02851.x>
3. Tenovuo J. Antimicrobial agents in saliva--protection for the whole body. *J Dent Res* 2002;81(12):807-9. <https://doi.org/10.1177/1544059102081012>
4. Barka T. Biologically active polypeptides in submandibular glands. *J Histochem Cytochem* 1980;28(8):836-59. <https://doi.org/10.1177/28.8.7003006>
5. Rougeot C, Messaoudi M, Hermitte V, Rigault AG, Blisnick T, Dugave C, et al. Sialorphan, a natural inhibitor of rat membrane-bound neutral endopeptidase that displays analgesic activity. *Proc Natl Acad Sci U S A*

- 2003;100(14):8549-54.  
<https://doi.org/10.1073/pnas.1431850100>
6. Isenman L, Liebow C, Rothman S. The endocrine secretion of mammalian digestive enzymes by exocrine glands. *Am J Physiol* 1999;276(2 Pt 1):E223-32.  
<https://doi.org/10.1152/ajpendo.1999.276.2.E223>
  7. Kongshavn PA, Lapp WS. Immunosuppressive effect of male mouse submandibular gland extracts on plaque-forming cells in mice: abolition by orchietomy. *Immunology* 1972;22(2):227-30.
  8. Kemp A, Mellow L, Sabbadini E. Suppression and enhancement of in vitro lymphocyte reactivity by factors in rat submandibular gland extracts. *Immunology* 1985;56(2):261-7.
  9. Zhao L, Lai Y, Jiao H, Huang J. Nerve growth factor receptor limits inflammation to promote remodeling and repair of osteoarthritic joints. *Nat Commun* 2024;15:3225. <https://doi.org/10.1038/s41467-024-47633-6>
  10. Tse DT, Latting MW, Hui JI. Epithelial tumors of the lacrimal gland. In: Albert D, Miller J, Azar D, Young LH, eds. *Albert and Jakobiec's Principles and Practice of Ophthalmology*. Springer, Cham 2020.  
[https://doi.org/10.1007/978-3-319-90495-5\\_68-1](https://doi.org/10.1007/978-3-319-90495-5_68-1)
  11. Vicente P, Marqués A, Aytés B, Escoda G. Minor salivary gland tumors: A clinicopathological study of 18 cases. *Med Oral Patol Oral Cir Bucal* 2008;13(9):E582-8.
  12. Ethunandan M, Davies B, Pratt CA, Puxeddu R, Brennan PA. Primary epithelial submandibular salivary gland tumors – Review of management in a district general hospital setting. *Oral Oncol* 2009;45(2):173-6.
  13. Joshi P, Bhattacharya K, Menon MB, Ghosh Laskar S, Joshi A, Patil V, et al. Malignant minor salivary gland neoplasms of larynx: Our experience. *Otolaryngol Pol* 2023;77(5):1-7.  
<https://doi.org/10.5604/01.3001.0053.4040>
  14. Sarmento DJ, Morais ML, Costa AL, Silveira EJ. Minor intraoral salivary gland tumors: a clinical-pathological study. *Einstein (Sao Paulo)* 2016;14(4):508-12.  
<https://doi.org/10.1590/S1679-45082016AO3749>
  15. Eveson JW, Cawson RA. Salivary gland tumors. A review of 2410 cases with particular reference to histological types, site, age and sex distribution. *J Pathol* 1985;146(1):51-8.  
<https://doi.org/10.1002/path.1711460106>
  16. Auclair PL, Ellis GL, Gnepp DR, Wenig BN, Janey CG. Salivary gland neoplasms: general considerations. In: Ellis GL, Auclair PL, Gnepp DR, eds. *Surgical pathology of the salivary glands*. Philadelphia: WB Saunders 1991:135-64.
  17. Guzzo M, Locati LD, Prott FJ, Gatta G, McGurk M, Licitra L. Major and minor salivary gland tumors. *Crit Rev Oncol Hematol* 2010;74(2):134-48.  
<https://doi.org/10.1016/j.critrevonc.2009.10.004>
  18. Alsanie I, Rajab S, Cottom H, Adegun O, Agarwal R, Jay A, et al. Distribution and frequency of salivary gland tumors: An international multicenter study. *Head Neck Pathol* 2022;16(4):1043-54.  
<https://doi.org/10.1007/s12105-022-01459-0>
  19. Yu GY, Ma DQ. Carcinoma of the salivary gland: a clinicopathologic study of 405 cases. *Semin Surg Oncol* 1987;3(4):240-4.  
<https://doi.org/10.1002/ssu.2980030405>
  20. Lakhani SR, O'Hare MJ. The mammary myoepithelial cell - Cinderella or ugly sister? *Breast Cancer Res* 2000;3:1. <https://doi.org/10.1186/bcr260>
  21. Shrestha S, Pandey G, Pun CB, Bhatta R, Shahi R. Histopathological pattern of salivary gland tumors. *J Pathol Nepal* 2014;4:520-4.
  22. Bashir S, Mustafa F, Malla HA, Khan AH, Rasool M, Sharma S. Histopathological spectrum of salivary gland tumors: A 10-year experience. *Sch J App Med Sci* 2013;1(6):1070-4.  
<https://doi.org/10.36347/sjams.2013.v01i06.0089>
  23. Cohen EG, Patel SG, Lin O, Boyle JO, Kraus DH, Singh B, et al. Fine-needle aspiration biopsy of salivary gland lesions in a selected patient population. *Arch Otolaryngol Head Neck Surg* 2004;130(6):773-8.  
<https://doi.org/10.1001/archotol.130.6.773>
  24. Chatterjee T, Panda PK. A pathological study of benign and malignant tumors of salivary glands. *Med J Armed Forces India* 2000;56(4):282-6.  
[https://doi.org/10.1016/S0377-1237\(17\)30208-3](https://doi.org/10.1016/S0377-1237(17)30208-3)

25. Bhargava S, Sant MS, Arora MM. Histomorphological spectrum of minor salivary gland tumors. *Indian J Cancer* 1982;19:134-40.
26. Shafkat Ahmed, Mohammad Lateef, Rouf Ahmad. Clinicopathological study of primary salivary gland tumors in Kashmir. *JK Practitioner* 2002;9(4):231-3.
27. Ito FA, Ito K, Vargas PA, de Almeida OP, Lopes MA. Salivary gland tumors in a Brazilian population: a retrospective study of 496 cases. *Int J Oral Maxillofac Surg* 2005;34(5):533-6.  
<https://doi.org/10.1016/j.ijom.2005.02.005>
28. Nagarkar NM, Bansal S, Dass A, Singhal SK, Mohan H. Salivary gland tumors - our experience. *Indian J Otolaryngol Head Neck Surg* 2004;56(1):31-4.  
<https://doi.org/10.1007/BF02968769>
29. Panda PK, Agarwal S. Tumors of salivary glands. *Indian J Cancer* 1962;6:82.
30. Khazanchi RK, Saba SS, Mittal D, Dhawan IK. Tumors of parotid glands-A review of 86 patients and current methods of treatment. *Indian J Cancer* 1988;25:1-6.
31. Soni D, Mathur K, Yadav A, Kumar V. Histopathological spectrum of salivary gland lesions in tertiary care centre at SMS Medical College, Jaipur, Rajasthan. *Int J Med Res Prof* 2016;2(2):209-15.
32. Pai RR, Sahu K, Kini AU. Clear cell predominant epithelial-myoepithelial carcinoma of the hard palate - role of IHC. *Indian J Otolaryngol Head Neck Surg* 2008;60(2):163-5. <https://doi.org/10.1007/s12070-008-0002-4>
33. Kim SH, Park SE, Bae HG, Song DH, Oh HH, Cho KR, et al. Epithelial-myoepithelial carcinoma of the nasopharynx: A case report and review of the literature. *Oncol Lett* 2015;10(2):927-30.  
<https://doi.org/10.3892/ol.2015.3314>
34. Seethala RR, Barnes EL, Hunt JL. Epithelial myoepithelial carcinoma: a review of the clinicopathologic spectrum and immunophenotypic characteristics in 61 tumors of salivary glands and upper aerodigestive tract. *Am J Surg Pathol* 2007;31(1):44-57.  
<https://doi.org/10.1097/01.pas.0000213314.74423.d8>
35. Chetty R. Intercalated duct hyperplasia: possible relationship to epithelial-myoepithelial carcinoma and hybrid tumors of salivary gland. *Histopathology* 2000;37(3):260-3. <https://doi.org/10.1046/j.1365-2559.2000.00976.x>