Journal of Research in Applied and Basic Medical Sciences 2023; 9(2): 87-96



## RABINS Journal of Research in Applied and Basic Medical Sciences

Original Article

# Effect of a significant deficiency in some antioxidants on semen parameters in cases of male infertility in Diyala Province, Iraq

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## Abstract

**Background & Aims:** Scientific evidence indicates that oxidative stress (OS) is a major factor in the etiology of male infertility. OS occurs when there is an imbalance between reactive oxygen species (ROS) and protective antioxidants. Current study aimed to evaluate some antioxidants and their effects on semen parameters in infertile Iraqi men.

*Materials & Methods*: In this study, 47 infertile males and 21 healthy people as the control group were enrolled. parameters for seminal fluid were including count, motility and morphology for assessment of semen quality. Glutathione, Vitamin E, Zinc and Selenium were also measured.

**Results:** Findings showed that sperm counts, percent of sperm motility and percent of normal morphology were significantly lower in the infertile male group compared to healthy group (p<0.001); whereas, abnormal motility and morphology in infertile male group were considerably higher than healthy group (p<0.001). Moreover, Results showed significant decrease in levels of Glutathione, Vitamin E, Zinc and Selenium in the infertile male group than the healthy group (p<0.001). Additionally, study reported a non-significant correlation between glutathione, vitamin E, zinc and selenium with semen parameters (P>0.05), except zinc level which showed significant correlation with the normal morphology of sperm (P<0.05). The results also showed a significant decrease in Vitamin E, Zinc and selenium levels in abnormal infertile groups compared to healthy group.

*Conclusion:* We believe that male infertility may be related with low antioxidant levels. Therefore, future studies should concentrate on enzymatic and non-enzymatic antioxidants, as well as genetic susceptibility and its effects on sperm quality.

Keywords: Glutathione, Male Infertility, Vitamin E, Selenium, Zinc

#### Received 07 December 2022; accepted for publication 27 March 2023

## Introduction

The inability to achieve spontaneous pregnancy after 12 months or more of regular unprotected sexual contact is known as infertility. Infertility affects couples on a variety of levels; recent studies showed that infertility is affecting about 15% of couples worldwide and male factor is responsible for almost 50% of infertility cases (1). According to the International Classification of Diseases, 11th Revision (ICD-11) (Geneva: WHO 2018) male infertility is frequently brought on by changes in semen ejection, a lack of sperm or sperm levels, or changes in sperm morphology and motility (2). Male subfertility's idiopathic causes have only recently been the subject of clinical and laboratory research efforts. The significant role of oxidative stress at the cellular level has been highlighted by researchers (3). It is estimated that half of all infertility cases between 25% and 87% of male subfertility is caused by the harmful effects of oxidative stress on sperm, and 1 in 20 man will be affected by subfertility (4). The overwhelming scientific evidence indicates that oxidative stress (OS) is a major factor in the etiology of male infertility (5). OS can occur when protective antioxidants and reactive oxygen species (ROS) are not balanced. OS can result in abnormal sperm parameters, significant sperm DNA damage, and even apoptosis. (6,7).

The term "male oxidative stress infertility" (MOSI) has recently been proposed, pointing out that between 30% and 80% of infertile men have elevated ROS in their sperm, affecting about 37.2 million infertile men (8). Exogenous or endogenous factors may be to blame for elevated ROS levels. The most frequent exogenous causes of oxidative stress in reproductive cells include obesity, poor nutrition, smoking, alcohol use, and environmental pollution. Additional causes recognized as endogenous are infections, autoimmune diseases, and chronic illnesses (9). Therefore, maintaining the redox potential balance, or balancing ROS by antioxidants, is necessary to maintain the optimal functioning of sperm cells (10).

Antioxidants are classified into two types: enzymatic antioxidant systems and non-enzymatic antioxidant systems. The enzymatic system consists of ascorbate peroxidase, alpha-dioxygenase, dehydroascorbate reductases, catalase, glutathione reductase, glutathione peroxidase, NADPH oxidase, Glutathione-S-transferase, superoxide dismutase, and peroxiredoxin. On the other hand, the non-enzymatic system consists of ascorbate, vitamin A, bilirubin, uric acid, glutathione, fibrin, mycothiol, melatonin, serum albumin, and phenolics, in addition to a variety of ingredients that are ingested along with food, including dietary supplements such as vitamins (11-14). Although a connection between systemic antioxidant deficiency and male infertility has not yet been documented, it's possible that some infertile men may be at the risk for antioxidant deficiency, especially those with certain lifestyles like smoking, drinking more alcohol, and dieting, who may also be at high risk for vitamin or antioxidant deficiencies (15). Nowadays, it is well known that a number of natural antioxidants, including alpha-lipoic acid, inositol, folate, zinc, selenium, coenzyme Q10, and vitamins are linked to an improvement in sperm quality by preventing OS (2). Due to the importance of the effect of antioxidants and their relationship to fertility, in our study, Glutathione, Vitamin E, Zinc and Selenium were chosen because of their important roles and significant effects in neutralizing the activity of free radicals and protecting sperm, as well as essential elements in the formation of sperm and testosterone biosynthesis (16). So, the present study aims to evaluate some antioxidants and effects on semen parameters in infertile Iraqi men.

## **Materials & Methods**

#### **Collection of seminal fluid:**

The semen samples of forty-seven male infertile participants in this study and twenty-five healthy persons who recorded as the control group was collected after at least three days without having sex. Semen samples were collected in a sterile, clean,

wide-mouthed, and labeled disposable plastic container by asking the groups to masturbate in the room next to the laboratory. Containers were closed and labeled (name, age, time of ejaculation, and duration of abstinence). To allow for liquefaction, the specimens were placed in an incubator at 37°C for 15-30 minutes. Following liquefaction and immediate processing, the following seminal fluid routine parameters were evaluated using the methods recommended by the World Health Organization (WHO) (17). After that, according to the guidelines of the WHO, the male group of infertile people were divided three categories: into azoospermia, oligozoospermia, and asthenozoospermia (18,19).

The parameters included sperm count, morphology, motility, viscosity, pH, and sperm appearance (18). Non-enzymatic antioxidant functions were evaluated using tests for Glutathione, Vitamin E, Zinc and Selenium after recording abnormal semen results. The participants' samples were taken between September 15, 2021, and August 20, 2022.

#### **Collection of samples:**

The samples were collected from different locations of Diyala Province, Iraq, from the healthy people as control group and infertile males as the case group from the auditors to consulting clinics and specialized laboratories in Baqubah City, Diyala Province, Iraq, for a purpose of diagnosis and treatment. The age ranged from 29 to 46 years.

After being informed about the study, each participant provided informed consent. As a result, the entire process of the investigation was based on the participants' agreed assent. In addition, all participants' weight and height were measured to calculate their Body Mass Index (BMI) using the equation = weight / (height)2 (20).

Six milliliters of venous blood were collected from the healthy and infertile groups and deposited in separate tubes. The serum samples were collected from the clotted blood using a centrifuge at 5000 rpm for 10 minutes. The serum was then stored in an eppendorf tube at -20°C. The enzyme linked immunosorbent assay (ELISA) was performed to evaluate the levels of Glutathione and Vitamin E (kits from Sunlong Biotech, China), whereas, Zinc and Selenium level were measured by atomic absorption.

#### Statistical analysis:

For the statistical analysis, SPSS (Statistical Package for Social Sciences) version 17.01 was used to examine statistical differences in parameter outcomes. In the study, an independent samples t-test was used to analyze the raw data and compare two groups' parameters; so there the mean and standard deviation were determined in study. Pearson's correlation test was used to investigate the relationship between non-enzymatic antioxidants and sperm parameters. Analysis of variance (ANOVA) were employed to compare non-enzymatic antioxidant levels in different groups. The threshold for statistical significance was set at a p-value of less than 0.01 or 0.05.

## Results

As shown in the table 1, healthy and infertile male groups were similar in terms of age with significant difference (p<0.05). However, there was no statistically significant difference between both groups, in the average BMI (p=0.212). Table 1 displays the mean values of the studied sperm parameters in the healthy and infertile groups. The healthy group had significantly higher levels of sperm count, normal motility, and the proportion of normal morphology than the infertile male group (p<0.001). However, only proportion of abnormal motility and abnormal morphology in infertile male group had significantly higher levels than healthy group had significantly higher levels than healthy group (p<0.001).

_	Infertile male group	Healthy group	– P value
Variables	Mean (SD)	Mean (SD)	
	n=47	n=25	
Age (years)	34.36 (4.1)	32.28 (2.1)	< 0.05*
BMI (kg/m <sup>2</sup> )	22.32 (3.6)	21.26 (2.9)	0.212 NS
Sperm counts (million/ml)	34.61 (10.9)	77.20 (12.6)	< 0.001**
Sperm motility (%)			
Normal	39.34 (10.2)	73.84 (10.7)	<0.001**
Abnormal	60.66 (10.2)	26.16 (10.7)	< 0.001**
morphology (%)			
Normal	48.20 (16.0)	81.32 (6.1)	< 0.001**
Abnormal	51.80 (16.0)	18.68 (6.1)	< 0.001**

Table 1: Parameters of semen quality in infertile males and the healthy group

NS: Non-significant, \*\* (P<0.01), \* (P<0.05)

According to the data in the table 2, serum nonenzymatic antioxidant levels were significantly lower in the infertile male group than in the healthy group. The results showed that serum glutathione levels in infertile male group were lower than in the healthy group (P<0.001). In addition, serum vitamin E levels were lower in the infertile male group than in the healthy group (P<0.001). Furthermore, both serum zinc and Selenium levels were lower in the infertile male group than the healthy group (P<0.001).

 Table 2: Comparison of levels of glutathione, vitamin E, zinc and selenium between infertile males and the healthy group

	Infertile male group	Healthy group	
Variables	Mean (SD)	Mean (SD)	- P value
	n=47	n=25	
Glutathione µmol/L	7.94 (1.1)	10.39 (0.9)	<0.001**
Vitamin E mg/L	4.77 (0.7)	5.88 (1.2)	<0.001**
Zinc µmol/L	9.10 (1.6)	11.42 (1.2)	<0.001**
Selenium µg/dL	4.30 (0.8)	4.84 (0.7)	<0.001**

\*\* (P<0.01)

Table 3 indicates that there was no significant correlation between serum glutathione and the parameters of semen quality in infertile male group. Moreover, there was no significant correlation between serum vitamin E and the parameters of semen quality in infertile male group. Additionally, no significant correlation between serum selenium and the parameters of semen quality in infertile male group. However, a significant correlation was found between serum zinc level and morphology of sperm. There was no correlation between serum zinc level and residual parameters of semen quality in infertile male group.

semen parameters for In	nertrie male group				
Variables	Infertile male gro	oup n=47			
	Glutathione	Vitamin E	Zinc	Selenium	

**Table 3:** correlation coefficient (r) values between serum glutathione, vitamin E, zinc and selenium levels with semen parameters for Infertile male group

Glutathione Vitamin E Zinc (r) (r) (r) (r) Sperm counts 0.086 -0.111 -0.243 -0.167 Normal Sperm motility (%) -0.012 0.105 -0.015 0.076 Normal morphology (%) 0.127 -0.113 -0.369\* 0.130

\* (P<0.05)

Table 4 indicates a significant decrease in serum glutathione levels, azoospermia, oligozoospermia and asthenozoospermia in infertile group compared to healthy group (P<0.05). There was a significant decrease in the serum levels of vitamin E of oligozoospermia, asthenozoospermia and Azoospermia groups compared to healthy group

(P<0.05). Additionally, there were also a significant decrease in serum zinc levels in oligozoospermia, asthenozoospermia and Azoospermia groups compared to healthy group (P<0.05). Furthermore, the result showed a significant decrease in serum selenium levels in oligozoospermia, asthenozoospermia, and azoospermia groups compared to healthy group (P<0.05).

**Table 4:** Comparison of levels of glutathione, vitamin E, zinc and selenium between abnormal infertile groups

 and the healthy group

groups	Variables			
	Glutathione µmol/L Mean (SD)	Vitamin E mg/L Mean (SD)	Zinc μmol/L Mean (SD)	Selenium µg/dL Mean (SD)
Healthy	10.39 (0.9) *	5.88 (1.2) *	11.42 (1.2) *	4.84 (0.7) *
Asthenozoospermia	8.08 (0.9)	4.75 (0.7)	8.94 (1.5)	4.26 (1.0)
Oligozoospermia	7.83 (1.2)	4.90 (0.6)	9.50 (1.8)	4.51 (0.5)
Azoospermia	7.71 (1.3)	4.57 (0.8)	8.80 (1.0)	4.01 (0.9)

\* (P<0.05)

## Discussion

According to the estimates, infertility problems impact about 25% of men globally. The process of spermatogenesis is impacted by a number of factors, including nutrition, genetics, physiological problems, and the environment (21). The current study indicated that ages of both groups were comparable (p<0.05) (table1), which is consistent with the findings of Alnajjar et al. (22). Most infertile patients were found to be between the ages of 25 and 50. The characteristics of sperm analysis revealed significant decrease in sperm count, sperm motility, and normal sperm morphology in the infertile male group as compared to the healthy group. Our findings concur with the results of Al-Baldawi et al. (23), who reported that sperm quality affects spermatozoa's ability to become fertilized. Furthermore, the findings of a study conducted by Al Jebory et al. (24) indicated that reduced consumption of some antioxidant agents in the group of infertile males is related to increased risk of poor sperm concentration, motility, and morphology. The current study showed a highly

significant decline in glutathione levels in infertile male group compared to healthy group counterparts. It has been established that a glutathione deficiency leads to spermatozoa instability which results in defective sperm motility. This implies that deficient glutathione levels may have an impact on male fertility (25). Moreover, increased levels of lipid peroxidation and free radical formation are lowering antioxidant levels in the blood as a result of continual oxidation of these antioxidants during the neutralization of these free radicals, showing that the antioxidant defense system is impaired in infertile males (26). Our study results showed significant decrease of vitamin E levels in infertile male group compared to healthy group counterpart, and this finding is consistent with the study of Rengaraj et al. (27), which showed that vitamin E deficiency impairs fertility in both humans and laboratory animals. Generally, found in cell membranes, vitamin E is regarded as a fat-soluble organic molecule that shields sperm cell membranes from oxidative damage (28). Any variation in vitamin E levels may have a role in infertility. Superoxide anion, hydrogen peroxide, and hydroxyl radicals are all immediately neutralized by vitamin E; hence an increase in these free radicals may cause vitamin E to become depleted (29). Additionally, our findings showed a significant decrease in zinc levels in the group of infertile males compared to their counterparts in the healthy group, and this finding is consistent with the results of Al-Baldawi et al. (23), who demonstrated the importance of zinc in fertility through its direct and indirect effects on spermatogenesis. Zinc plays important roles in the early phases of germ cell development and spermatogenesis, sperm cell maturation and development, ejaculation, liquefaction, and fertilization (30). As a result, zinc deficiency may be a major risk factor for poor sperm quality and idiopathic male infertility (31). Regarding the results of the study, there was a significant decrease in selenium levels in infertile male group compared to healthy group counterparts. Selenium appears to be important for sperm cells' antioxidative defense.

Spermatogenesis is significantly impacted when a breakdown in selenium homeostasis occurs (32).

The results of our study agreed with the study of Onono et al. (33) which showed that selenium deficiency in men leads to oligospermia and hence impaired spermatogenesis, decreased sperm motility and ability to fertilize, and an increase in the amount of aberrant male sex cells as a result of oxidative stress damages on integrity of their membranes. In the current study, we report a non-significant correlation between glutathione, vitamin E and selenium with semen parameters, as well as, a non-significant correlation was observed between zinc and sperm counts and motility; serum zinc levels showed significant correlation with normal morphology of sperm. Similar results have been reported in earlier researches (34,35).

There is evidence that zinc influences spermatozoa's physiological processes such as their motility and morphology, and a decline in seminal zinc content results in poor-quality semen and fewer opportunities for fertilization (36). Additionally, though the reasons for the decline in male fertility are still unknown, a number of lifestyle factors, particularly an unhealthy diet, are thought to be major causes of male reproductive health impairment. However, roughly 30% of cases of male infertility are known as idiopathic infertility since they continue to have unidentified reasons (37,38). In the present study, we report a significant decrease in serum glutathione levels in azoospermia, oligozoospermia and asthenozoospermia groups compared to healthy group. This result agree with Krzyściak et al. (39) which indicated a decrease in glutathione in the abnormal infertile groups compared to healthy group. Numerous studies have reported a decrease in glutathione concentration in the groups of oligozoospermia and asthenozoospermia, which may be related to a decrease in NDPH coenzyme. NDPH is necessary for the production of glutathione and also is a catalytic agent for the enzyme glutathione reductase, which works to convert glutathione from its GSSG (glutathione disulfide) form to glutathione (40,41). On

the other hand, results of the study showed a significant decrease in serum zinc, vitamin E, and selenium levels in oligozoospermia, asthenozoospermia and azoospermia groups compared to healthy group. In summary, several studies have shown deficiency effects of zinc, selenium and vitamin E in men with infertility. A number of studies showed that insufficient zinc levels prevent spermatogenesis, contribute to sperm abnormalities, and diminish blood testosterone levels (42).

In contrast, selenium seems to have a favorable effect on the Leydig cell, and thus in turn, affects the release of testosterone (32). Vitamin E is however an essential substance for preventing oxidative damage to cell membranes by capturing and scavenging free radicals within cellular membranes (43,44). Another study indicated that zinc, selenium, and glutathione correlations with indices of sperm quality and suggested that a decrease in seminal antioxidants may be a risk factor for sperm abnormalities and idiopathic male infertility (45). According to Ritchie et al. (46), idiopathic male infertility may be caused by decreased seminal plasma antioxidant activity and increased ROS generation. Studies showed that antioxidant therapy had a significant positive impact on basic sperm parameters, advanced sperm function, outcomes of assisted reproductive therapy, and live birth rate in a comprehensive review of the impact of oral antioxidants on male infertility. The most frequently used nutrients were vitamin C, vitamin E, N-acetyl cysteine, carnitines, coenzyme Q10, selenium, zinc, lycopene, and folic acid (47).

## Conclusion

In conclusion, the present study shows significant decrease in the serum levels of glutathione, vitamin E, zinc and selenium in infertile male group than in the healthy group. Thus, we believe that male infertility may be related with a low antioxidant levels. Future studies should concentrate on enzymatic and nonenzymatic antioxidants, as well as genetic susceptibility and its effects on sperm quality.

## Acknowledgments

The author would like to express and thank the doctors and nurses for their invaluable assistance during this study. The author wishes all of the infertile couples who participated in the study good health in their personal life.

#### **Ethical approval**

The study was approved by the Council College of Education for Pure Sciences/University of Diyala's local ethics committee (Code: CEPS.UD.REC NO.190.16/11/2023).

## References

 Mannucci A, Argento FR, Fini E, Coccia ME, Taddei N, Becatti M, et al. The impact of oxidative stress in male infertility. Front Mol Biosci 2021;8:799294. Available from:

http://dx.doi.org/10.3389/fmolb.2021.799294.

- Cilio S, Rienzo M, Villano G, Mirto BF, Giampaglia G, Capone F, Crocetto F. Beneficial effects of antioxidants in male infertility management: A narrative review. Oxygen 2022;2(1):1–11.
- Evans EPP, Scholten JTM, Mzyk A, Reyes-San-Martin C, Llumbet AE, Hamoh T, et al. Male subfertility and oxidative stress. Redox Biol 2021;46(102071):102071. Available from:

http://dx.doi.org/10.1016/j.redox.2021.102071.

- de Ligny W, Smits RM, Mackenzie-Proctor R, Jordan V, Fleischer K, de Bruin JP, et al. Antioxidants for male subfertility. Cochrane Database Syst Rev 2022;5(5):CD007411. Available from: http://dx.doi.org/10.1002/14651858.CD007411.pub5.
- Russo A, Troncoso N, Sanchez F, Garbarino J, Vanella A. Propolis protects human spermatozoa from DNA damage caused by benzo [a] pyrene and exogenous reactive oxygen species. Life Sci 2006;78(13):1401-6.
- Li KP, Yang XS, Wu T. The effect of antioxidants on sperm quality parameters and pregnancy rates for idiopathic male infertility: A network meta-analysis of randomized controlled trials. Front Endocrinol (Lausanne) 2022;13:810242. Available from: http://dx.doi.org/10.3389/fendo.2022.810242.

- Celik G, Günaydin G, Demir B, Yilmaz F, Ersen T, Tuncay M, et al. Oxidative stress in patients with carbon monoxide poisoning. Ankara Med J 2021;23(1).
- Agarwal A, Parekh N, Panner Selvam MK, Henkel R, Shah R, Homa ST, et al. Male Oxidative Stress Infertility (MOSI): Proposed terminology and clinical practice guidelines for management of idiopathic male infertility. World J Mens Health 2019;37(3):296–312. Available from:

http://dx.doi.org/10.5534/wjmh.190055.

- Tremellen K. Oxidative stress and male infertility—a clinical perspective. Hum Reprod Update. 2008;14(3):243-58.
- Kowalczyk A. The role of the natural antioxidant mechanism in sperm cells. Reprod Sci 2022;29(5):1387–94. Available from: http://dx.doi.org/10.1007/s43032-021-00795-w.
- Lu Z, Wen T, Wang Y, Kan W, Xun G. Peripheral nonenzymatic antioxidants in patients with schizophrenia: a case-control study. BMC Psychiatry 2020;20(1):241. Available from: http://dx.doi.org/10.1186/s12888-020-02635-8.
- Haida Z, Hakiman M. A comprehensive review on the determination of enzymatic assay and nonenzymatic antioxidant activities. Food Sci Nutr 2019;7(5):1555– 63. Available from:

http://dx.doi.org/10.1002/fsn3.1012.

- Irato P, Santovito G. Enzymatic and non-enzymatic molecules with antioxidant function. Antioxidants (Basel) 2021;10(4):579. Available from: http://dx.doi.org/10.3390/antiox10040579.
- Yazici G, GÜRGEN S, SUNAR M, ELMAS Ç. Effects of Various Antioxidants on Rat Lung Tissue During Chemotherapy: Electron Microscopic Study. Gazi Med J 2021;32(3).
- Zini A, Al-Hathal N. Antioxidant therapy in male infertility: fact or fiction? Asian J Androl 2011;13(3):374–81. Available from: http://dx.doi.org/10.1038/aja.2010.182.
- Ahmadi S, Bashiri R, Ghadiri-Anari A, Nadjarzadeh A. Antioxidant supplements and semen parameters: An evidence based review. Int J Reprod Biomed 2016;14(12):729-736.

- World Health O. WHO Reference values of semen variables. In: Laboratory Manual for the Examination of Human Semen and Sperm- Cervical Mucus Interaction. 4th. Ed. Cambridge University Press. Cambridge;1999.
- World Health O. WHO Laboratory manual for the examination of human semen and sperm cervical mucus interaction. Cambridge university press; 2009.
- World Health O. WHO laboratory manual for the examination and processing of human semen. 5th ed. Cambridge University press, Cambridge. United States. Geneva: World Health Organization; 2010.
- 20. Nuttall FQ. Body mass index: Obesity, BMI, and health: A critical review. Nutr Today 2015;50(3):117–28. Available from: http://dx.doi.org/10.1097/NT.00000000000092.
- Alkumait M, Abdul-Aziz M, Nima M. The effect of glutathione versus co-enzyme Q10 on male infertility original study. Medico Leg Update 2020;20:409-14
- Alnajjar AF, Qahraman AS, Ismael AA, Hamza TA Karim YS Al-Dhalimy AMB, Prevalence of Male Infertility In Kirkuk City, Iraq. J Pharm Negative Results 2022;13(1):89-94.
- Al-Baldawi AT, Naji NA, Al-Ani AA. Male infertility and physiological role of zinc. Iraqi J Med Sci 2000:67.
- Al Jebory KH, Al Mukhtar SH. Relationship between dietary intake of antioxidant (vitamins C, E, and selenium) with semen quality. Mosul J Nurs 2015;3(1):1-5.
- Naher ZU, Biswas SK, Mollah FH, Ali M, Arslan MI. Role of glutathione in male infertility. Banglad J Med Biochem 2013;4(2):20–5. Available from: http://dx.doi.org/10.3329/bjmb.v4i2.13772.
- 26. Naif W, Al-Salih R, Guzar S. Assessment of lipid peroxidation and glutathione levels in serum and seminal plasma for unexplained infertile men in Thiqar governorate. J College Edu Pure Sci 2018;8(3):57– 50. Available from:

http://dx.doi.org/10.32792/utq.jceps.08.03.06.

 Rengaraj D, Hong YH. Effects of dietary vitamin E on fertility functions in poultry species. Int J Mol Sci 2015;16(5):9910–21. Available from: http://dx.doi.org/10.3390/ijms16059910.  Walczak-Jedrzejowska R, Wolski JK, Slowikowska-Hilczer J. The role of oxidative stress and antioxidants in male fertility. Cent European J Urol 2013;66(1):60– 7. Available from:

http://dx.doi.org/10.5173/ceju.2013.01.art19.

- Mukhtar NJA, Temimmi HMA, Edan BJ. Local and Systemic Free Radicals level in Unexplained Infertile Women. Kufa Med J 2012;15(1).
- 30. Vickram S, Rohini K, Srinivasan S, Nancy Veenakumari D, Archana K, Anbarasu K, et al. Role of zinc (Zn) in human reproduction: A journey from initial spermatogenesis to childbirth. Int J Mol Sci 2021;22(4):2188. Available from:

http://dx.doi.org/10.3390/ijms22042188.

- 31. Fatima P, Hossain MM, Rahman D, Mugni CR, Hossain HB, Hossain HN, Sumon GM. Impact of Seminal Plasma Zinc and Serum Zinc Level on Semen Parameter of Fertile and Infertile Males. J Bangladesh Coll Phys Surg 2017;35(1):15-9.
- 32. Akinloye O, Arowojolu AO, Shittu OB, Adejuwon CA, Osotimehin B. Selenium status of idiopathic infertile Nigerian males. Biol Trace Elem Res 2005;104(1):9– 18. Available from:

http://dx.doi.org/10.1385/BTER:104:1:009.

- 33. Bizerea-Moga TO, Pitulice L, Bizerea-Spiridon O, Moga TV. Evaluation of serum selenium status by age and gender: A retrospective observational cohort study in western Romania. Nutrients 2021;13(5):1497.
- 34. Onono L, Orinda G, Munga L. Evaluation of zinc in blood, semen and their relationship to sperm quality among males attending infertility clinic in Kenyatta National Hospital. East Afr Med J 2021;98(10):4236-45.
- 35. Kothari RP, Chaudhari AR. Zinc levels in seminal fluid in infertile males and its relation with serum free testosterone. J Clin Diagn Res 2016;10(5):CC05-8. Available from:

http://dx.doi.org/10.7860/JCDR/2016/14393.7723.

36. Osadchuk L, Kleshchev M, Danilenko A, Osadchuk A. Impact of seminal and serum zinc on semen quality and hormonal status: A population-based cohort study of Russian young men. J Trace Elem Med Biol 2021;68(126855):126855. Available from: http://dx.doi.org/10.1016/j.jtemb.2021.126855.

37. Salas-Huetos A, James ER, Aston KI, Jenkins TG, Carrell DT. Diet and sperm quality: Nutrients, foods and dietary patterns. Reprod Biol 2019;19(3):219–24. Available from:

http://dx.doi.org/10.1016/j.repbio.2019.07.005.

- 38. Jungwirth A, Giwercman A, Tournaye H, Diemer T, Kopa Z, Dohle G, et al. European Association of Urology guidelines on Male Infertility: the 2012 update. Eur Urol 2012;62(2):324–32. Available from: http://dx.doi.org/10.1016/j.eururo.2012.04.048.
- 39. Krzyściak W, Papież M, Bąk E, Morava E, Krzyściak P, Ligęzka A, et al. Sperm antioxidant biomarkers and their correlation with clinical condition and lifestyle with regard to male reproductive potential. J Clin Med 2020;9(6):1785. Available from: http://dx.doi.org/10.3390/jcm9061785.
- 40. Al Fleafil SJ, Al Faisal AHM, Mahood RA. Association between GSTM1, GSTT1 Genes Variants and Some Physiological Parameters in Infertility Patients. Iraqi J Biotech 2021;1(20).
- Wu G, Fang YZ, Yang S, Lupton JR, Turner ND. Glutathione metabolism and its implications for health. J Nutr 2004;134(3):489–92. Available from: http://dx.doi.org/10.1093/jn/134.3.489.
- 42. Fallah A, Mohammad-Hasani A, Colagar AH. Zinc is an essential element for male fertility: A review of Zn roles in men's health, germination, sperm quality, and fertilization. J Reprod Infertil 2018;19(2):69–81.
- Abbood MRJ. The Effect of Selenium and Vitamin E on Male Infertility. Med J Babylon 2012;9(1).
- 44. Vakili S, Zal F, Mostafavipour Z, Savardashtaki A, Jafari K Majid, Hassanpour A. Effects of quercetin and vitamin E on ovariectomy-induced oxidative stress in rat serum and tibia. Arch Biol Sci 2020;72(1):95–104. Available from: http://dx.doi.org/10.2298/abs191115003v.
- 45. Atig F, Raffa M, Habib B-A, Kerkeni A, Saad A, Ajina M. Impact of seminal trace element and glutathione levels on semen quality of Tunisian infertile men. BMC Urology 2012;12(1):1-8.

- Ritchie C, Ko EY. Oxidative stress in the pathophysiology of male infertility. Andrologia 2021;53(1):e13581.
- 47. Majzoub A, Agarwal A. Systematic review of antioxidant types and doses in male infertility:

Benefits on semen parameters, advanced sperm function, assisted reproduction and live-birth rate. Arab J Urol 2018;16(1):113–24. Available from: http://dx.doi.org/10.1016/j.aju.2017.11.013.

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